

Screening of Indigenous Bacteria in Tilapia (*Oreochromis* sp) as a Probiotic Candidate.

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Abstract: *Tilapia (Oreochromis sp.) are ranks the seventh largest freshwater species production in the world. Tilapia feed have the ability to digest feed protein only 75% and protein efficiency ratio of 23,5%. The use of high-protein diet will increase the production cost, therefore it will result in lower production profit. To increase protein efficiency ratio and digestibility by using indigenous bacteria originated from fish intestine can be an alternative way. The purpose this study was to isolate the indigenous bacteria in Tilapia as a potential probiotic candidate. Bacterial screening was conducted through several tests, i.e. pathogenicity, antagonism, characteristic of extracellular enzyme, growth characteristic and biochemical test. Pure culture of fourteen (14) isolates were isolated. All isolates were safe for fish as they have γ haemolytic effects on blood agar. Three isolates (NL 004, NL 007 and NL 013) were identified to have higher antagonism activities. Isolate NL 004 had the biggest capability for hydrolyzing protein, fat, and carbohydrate, (12,48 mm, 5,75 mm, 5,37 mm) respectively. Isolate NL 004 had the value best of growth rate (0,897 generation.hours⁻¹) and generation time (0,768 hours). Taking all the screening test resulted the best probiotic candidate NL 004 was a *Bacillus* sp.*

Key words : *Bacillus* sp, Feed efficiency, Isolation.

“1. Introduction”

Feed is the primary factor causing of growth increase. Commercial feed suck up 60-70% of production costs. Tilapia (*Oreochromis* sp.) require feed protein 28-40%, yet the tilapia only have protein efficiency ratio of 20-25% and the digestibility 75%. [1-3]. To improve of protein efficiency ratio and digestibility to do by increasing the number of bacterial digestion. The addition of probiotics indigen whit extraselluler digestive enzymes such as amylase, protease and lipase can be alternative way.

Probiotics are beneficial bacteria capable of producing various digestive enzymes can improve growth, feed efficiency, digestibility, increase resistance to bacterial pathogens and improve water

quality [3-6]. Probiotic bacteria could be isolated from the intestines of fish, so can be an indicator found the bacteria are able adhesive to the cell wall of fish intestines. The purpose of this study is to isolate the tilapia indigenous bacteria as a potential candidate probiotic, which should increase the digestibility of the feed.

“3. Materials and Methods”

2.1 Tilapia (*Oreochromis* sp)

Tilapia used around with an average weight of 9±0.21 g. From the pond at the central of Aquaculture Punten Malang east Java. Then the fish is cut and taken away part of the intestine using sectio set.

3.2 Isolations

Tilapia taken part of gut after it grounded, then made up a 10⁻⁸ dilution, dilution planted on MRSA sterile media, as many as 1 ml pour plate, and incubated in a temperature of 35°C for 48 hours. The number of colonies counted and do calcification (color, edges, surfaces and shape of colonies). To obtain pure cultures each colony is selected and then planted using ose with quadrant method, then incubation for 48 hours at a temperature of 35°C [7].

1.3 Pathogenicity test

The pathogenicity test is to determinate characteristic of the isolated bacteria. The test was done using *sheep blood agar* media whit by planting or scraping. The test results are pathogenic if there clear zone around the colony dan contrary [8].

1.4 Antagonism *in vitro* test

Antagonism test performed to determine the ability of bacteria isolation to inhibits the growth of pathogenic bacteria (*Aeromonas hydrophila* and *Edwardsiella tarda*). The test was done in a way planting pathogens MRSA media, and then paper disc placed in the middle of the media. After that

planting as many as 20 μL of bacterial isolation with a density of 10^8cfu.ml^{-1} . Positive results demonstrated the clear zone around the paper disc [6].

1.5 Extracellular enzyme test

The test performed to determine the the type of enzymes produced by candidate bacteria. The test was done by making modified test medium. In the test of enzyme with plate agar counte (PCA) whit much as 1% starch gr.V^{-1} , the test of lipase enzyme whit nutrient agar media (NA) add 1% gr.V^{-1} fish oil, and the test of protease enzyme whit backto agar (BA) media add 10% gr. V^{-1} casein. A positive result is shown on a clear zone around the paper disc [9].

1.6 Growth characteristics test

Isolation of bacteria taken as one ose then planted in sterile TSB media. Bacterial growth was observed using a spectrophotometer (Genesys 20) at 600 nm. Isolates was observed growth every hour on the first five hours, and after this whit two hous until for twenty nine hours. The results are compared with the pathogenic bacteria *A. hydrophila* and *E. tarda* [10].

1.7 Identification of bacteria

The identification test is morphological and biochemical. The morphological test is Gram stain and the biochemistry test include TSIA, GAS, H_2S , catalase, oxidase test and OF. The test derivatives of carbohydrates such as glucose, lactose, sucrose, maltose, mannitol, dulcitol, inositol, arabinose, raffinosa, trehalose, and xylose. The biochemistry synthesis test includes Gelatin test, motility, Indol, Simons citrate, Ornithine Decarboxylase, phenylalanine. The test results then matched to book Cowan And Steel's Manual For The Identification Of Medical Bacteria 3rd Edition. (Cowan, 1985).

“3. Results and Discussion”

The number of colonies that grow as much as of 117 colonies in the MRSA media overall. The number of colonies selected by color, edges, surfaces and shape of colonies then obtained 14 isolates were named (NL NL 001-014). The results of isolation and then test pathogenicity using sheep blood agar (Table 1). Pathogenicity test (test of haemolins) is necessary to see if the bacteria harmful to fish or host, pathogenic bacteria will produce exotoksin called haemolins that would

damage blood cells making arise in the clear zone around the colony [11].

Table 1. Test of Pathogenicity and Antagonisme

Isolate	Pathogenicity test			Antagonism (mm)	
	α	β	γ	<i>A. hydrophila</i>	<i>E. tarda</i>
NL 001	Nd	Nd	+	Nd	Nd
NL 002	Nd	Nd	+	Nd	Nd
NL 003	Nd	Nd	+	Nd	Nd
NL 004	Nd	Nd	+	15,75	16,98
NL 005	Nd	Nd	+	Nd	Nd
NL 006	Nd	Nd	+	Nd	Nd
NL 007	Nd	Nd	+	14,52	16,58
NL 008	Nd	Nd	+	Nd	Nd
NL 009	Nd	Nd	+	5,16	5,93
NL 010	Nd	Nd	+	Nd	Nd
NL 011	Nd	Nd	+	Nd	Nd
NL 012	Nd	Nd	+	Nd	Nd
NL 013	Nd	Nd	+	9,43	5,92
NL 014	Nd	Nd	+	Nd	Nd

Information : Nd = No detected

+ = Detected

The results test of antagonism bacterial candidates with pathogenic bacteria *A. hydrophila* and *E. tarda* **performed** (Table 1). Of the fourteen isolates, only four isolates have inhibitory effects. Only three highest isolate which is then used for next screening step. Antagonism test is necessary in to determine the ability of bacteria inhibit the growth of pathogenic bacteria with the ability to better grow [12]. The ability of antagonism is the natural activity that bacteria to secrete bacteriocins, lysozyme, and H_2O_2 [8].

Bacteriocins are proteins synthesized by ribosomes that are antagonistic to the organisme are on still family. Lysozyme damage the cell walls of bacteria by breaking the connection glycosides on the membrane peptidoglycan [13]. Produces enzymes myeloperoksida very important to used utilize oxygen free radicals to produce hypochlorous to kill pathogens [13]. Power of substance antagonist occurs with microbial agents that undermine and inhibit growth. Phagocytosis activity responded by first activating imflamantori response before the production of antibodies, and mediated by fagosisosis cells such as neutrophils, monocytes, and magrofag [14].

Characterization of the ability of bacteria to produce enzymes exogenous on selective media (Table 2). The three isolates obtained that able to produce proteolytic enzymes, amylyolytic and

lipolytic. The ability to produce the enzyme ekstraselluler will improve feed efficiency ratio and digestibility [6].

Table 2. The extracellular enzyme test isolates probiotic candidates.

Isolate	Diameter of clear zone (mm)		
	Protease	Lipase	Amilase
NL 004	12,48±0,67	5,75±0,146	5,37±0,026
NL 007	11,08±0,04	4,303±0,05	5,27±0,020
NL 013	11,53±0,06	3,78±0,166	4,35±0,045

☒ Digestive tract (gut) is rich in nutrients that is a favorite of some bacteria to grow, the abundance of intestinal bacteria will increase the production of digestive enzymes. Groups of bacteria such as *Bacillus* secretly issued a hydrolysis enzyme to degrade many substrates and to grow on a wide scale source of nutrients. Extracellular enzymes can increase the amount of protein of 20-25 g.L⁻¹ [15]. *Bacillus* will produce hydrolysis enzyme ektraselular (polysaccharide) [17].

Table 3. Characterization of bacterial growth for 29 hours.

Isolate	Karakter Uji		
	Specific growth rate (Generation.hour ⁻¹)	lag Fase (h)	Generation time (h)
NL 004	0,897	3	0,768
NL 007	0,5337	1	1,317
NL 013	0,629	1	1,09
A. <i>hydrop hila</i>	0,252	1	2,731
E. <i>tarda</i>	0,323	1	2,13

Characterization of the best growth rate was NL 004 0,897 Generations.hours⁻¹. Diffent of growth characterization affected the ability to utiliza nutrients (carbon), under culture conditions. Temperatute optimum of bacteria growth at 30 °C, whit a pH of 6-7 [18]. Stability of temperature will can be affect the activity sensitively of enzymes and growth [19]. Bacteria will be dormant because of environmental stress such as low temperature, nutrients disappear suddenly, and osmotic pressure [20]. Intestinal bacteria more resistant to low pH. The existence of sugar in the culture medium significantly affect (Desai 2008; Zhou *et al.*, 2013).

Morphology identification (Table 4) whit gram staining and then observed under a microscope with a magnification of 1000X. the fuction of gram staining to determine type of

bacteria by gram. The ability of baterial cells to absorb colors is different gram-positive absorbing of purple color, gram-negative while absorbing red. This reaction is caused by differences of cell wall.

In gram-negative contained lipid. whereas in gram-positive there its'n are so crystal violet can be through the cell wall and tied so purple, besides gram-positive have a layered peptidoglycan [21, 22]. Biochemical test results showed a positive reaction to the test Voges-Proskauer (VP), catalase, starch, citrate, nitrate, glucose, fructose, maltose, sucrose, ribose and trehalose [6]. Catalase test is used to determine the ability and enzymes produced by bacteria. TSIA test to determine the ability of a bacterium to ferment sugars to produce acids or gas [23]. The results were analyzed by using the book identification 3ndEdition Cowan and Steel (1985).

☒ Bacteria NL 004 unidentified gram + such findings in accordance with identification keys Cowan (1985), gram + bacteria with endospores, to shape of long rods (bacil) able to decipher H₂O₃ 3%, motile, reactive Aesculin, Simmons' citrate +, Glycerol +, are Aerobic and facultative anaerobic is the genus *Bacillus*, and *Clostridium*. Whereas if it is round (cocci) belong to the genus *Micrococcus* and *Staphylococu* [24]. The characteristics of the genus *Bacillus* are gram +, catalase +, fermentatif. *Bacillus* in action with flagella peritrichous, capable of hydrolyzing starch, gelatin, casein, to reduce nitrate and the V/P reaction is + [25,26]. From the key identification has been isolate NL 004 included in the types of bacteria *Bacillus*. *Bacillus* has been long used as a probiotic efficient because it can produce extracellular enzymes and anti-microbial components [6]. Some studies explain that *Bacillus* sp can be used as probiotics because it is more resistant to changes in the environment, can be combined with other beneficial bacteria and can be used as a biocontrol microbial pathogens in fish [13].

Table 4. Identification test result isolat NL 004

Num	Types of test	Isolate
		NL 004
1	Gram identification	
	Color colonies	Whit milk
	Edge of colonies	undulate
	Type of Gram	+
	Bacteria shape	Bacil
2	Biochemistry test	
	TSI Agar	A/A
	Gas	-
	H ₂ S (Sulfur acid)	-
	Catalase	+
	Oksidase	+
	O/F	Fermentatif

3	Fermentation of carbohydrate	
	Glucose	+
	Lactose	+
	Sucrose	+
	Maltose	+
	Mannitol	+
	Dulcitol	-
	Salicin	+
	Inositol	-
	Sorbitol	+
	Arabinose	+
	Raffinose	-
	Trehalose	+
	Xylose	+
4	Synthetic Biochemistry test	
	Gelatin	-
	Motility	+
	Indol	-
	Simmons Citrate	-
	Malonate	-
	Christensen's Urease	+
	Methyl Red (MR)	+
	Voges Proskauer (VP)	+
	Arginin Dihidrolase	+
	Lysine Decarboxylase	-
	Ornithin Decarboxylase	-
	Phenylalanin Deaminase	-
	Aesculin Hydrolisis	+
5	Result of analisis	<i>Bacillus</i> sp

Information: - (no reaction) ,+ (reaction), A/A (Acid/Acid).

The supplementation probiotic of *Bacillus* for 8 weeks can improve total leukocytes, phagocytosis activity, lysozyme activity, and serum super oxide dismutase the function to cut and inhibit xanthine and xanthine oxidase [27]. *Bacillus* increasing the immune response and increase resistance to vibrio, increases resistance to stress, improve survival and growth [3,4,28].

“4. Conclusion”

Based on the screening test pathogenicity, antagonism, extracellular enzymes and growth characteristic was found that the best candidates are NL 004 and in morphology and biochemistry test results are the type of *Bacillus* sp.

“5. Acknowledgements”

Thank you to all the parties concerned, in particular the research group of aquatic biofloc.

“6. References”

- [1] Maina JG, Beames RM, Higgs D, Mbugua PN, Iwama G, & Kisia SM. 2002. Digestibility and feeding value of some feed ingredients fed to tilapia *Oreochromis niloticus* (L.). *Aquaculture Research* 33: 853–62.
- [2] Abd El-Rhman AM, Khattab YAE, Shalaby AME. 2009. *Micrococcus luteus* and *Pseudomonas* species as probiotics for promoting the growth performance and health of Nile tilapia, *Oreochromis niloticus*. *Fish and Shellfish Immunology* 27: 175-180.
- [3] Telli GS, Maria JTRP, Danielle de CD, Fabio RS, Carlos MI, Leonardo T. 2014. Dietary administration of *Bacillus subtilis* on hematology and non-specific immunity of Nile tilapia *Oreochromis niloticus* raised at different stocking densities. *Fish and Shellfish Immunology* 39: 305-311.
- [4] Ayyat MS, HM Labib, HK Mahmoud. 2014. A Probiotic Cocktail as a Growth Promoter in Nile Tilapia (*Oreochromis niloticus*). *Journal of Applied Aquaculture* 263: .208–215.
- [5] Dash G, Ram PR, K Pani P, M Makesh, MA Pradee. 2014. Evaluation of *Lactobacillus plantarum* as feed supplement on host associated micro flora, growth, feed efficiency, carcass biochemical composition and immune response of giant freshwater prawn, *Macrobrachium rosenbergii* (de Man, 1879). *Aquaculture* 432: 225–236.
- [6] NavinChandran M, Palanisamy I, Subramanian M, Ramasamy R, Santhiyagu P, Grasian I, Arunachalam P. 2014. Influence of probiotic bacterium *Bacillus cereus* isolated from the gut of wild shrimp *Penaeus monodon* in turn as a potent growth promoter and immune enhancer in *P. monodon*. *Fish and Shellfish Immunology* 36: 38-45.
- [7] Klimentová J. 2015. Methods of isolation and purification of outer membrane vesicles from gram-negative bacteria. *Microbiological Research* 170: 1–9.
- [8] Apun-Molina JP, Apolinar SM, Antonio LG, Sergio FMD, Maurilia RC. 2009. Effect of potential probiotic bacteria on growth and survival of tilapia *Oreochromis niloticus* L., cultured in the laboratory under high density and suboptimum temperature. *Aquaculture Research* 40: 887-894.
- [9] Badriyah BI, T Ardyati, S Widyarti. 2014. Protease activity of bacterial isolates TP5K1 and TP6K5 in tofu solid waste substrate and identification of isolates based on 16S rDNA. *Int. J. Biosci* 5: 128-134.
- [10] Yuniarti A, DA Guntoro, AM Hariati. 2013. Response of Indigenous *Bacillus megaterium* Supplementation on the Growth of *Litopenaeus vannamei* (Boone), a New Target Species for Shrimp Culture in East Java of Indonesia. *Journal of Basic and Applied Scientific Research* 3: 747–754.
- [11] Egwuatu TO, Folasade TO, Isi MO, Bamiro J,

Damilola GA, OA Osinupebi. 2014. Effect of blood agar from different animal blood on growth rates and morphology of common pathogenic bacteria. *Advances in Microbiology* 4: 1237–1241.

[12] Klinkenberg, G. et al., 2010. Selection of candidate probiotics by two different screening strategies from Atlantic cod (*Gadus morhua* L.) larvae. , 144, pp.153–159.

[13] Gupta A, P Gupta, A Dhawan. 2014. Dietary supplementation of probiotics affects growth, immune response and disease resistance of *Cyprinus carpio* fry. *Fish and Shellfish Immunology* 41: 113-119.

[14] Sun Y, Hong-Ling Y, Ru-Long M, Wen-Yan L. 2010. Probiotic applications of two dominant gut *Bacillus* strains with antagonistic activity improved the growth performance and immune responses of grouper *Epinephelus coioides*. *Fish and Shellfish Immunology* 29: 803–809.

[15] Parrado J, B Rodriguez-Morgado, M. Tejada, T Hernandez, C Garcia. 2014. Enzyme and Microbial Technology Proteomic analysis of enzyme production by *Bacillus licheniformis* using different feather wastes as the sole fermentation media. *Enzyme and Microbial Technology* 57: 1–7.

[17] Balasubramanian N, N Simões. 2014. *Bacillus pumilus* S124A carboxymethyl cellulase ; a thermo stable enzyme with a wide substrate spectrum utility. *International Journal of Biological Macromolecules* 67: 132–139.

[18] Xu J, Hongjie W, Ziwei Z, Fang J, Xianchao Y, Qing H, Jianrong S. 2016. Isolation and characterization of *Bacillus amyloliquefaciens* ZDS-1 : Exploring the degradation of Zearalenone by *Bacillus* spp . *Food Control* 68: 244–250.

[19] Zhang H, Xiuzhen G, Jie R, Jinhui F, Tongcun Z, Qiaqing W, Dunming Z. 2014. Enzymatic hydrogenation of diverse activated alkenes. Identification of two *Bacillus* old yellow enzymes with broad substrate profiles. “*Journal of Molecular Catalysis. B, Enzymatic*”, 105: 118–125.

[20] Rahman MH, S Suzuki, K Kawai. 2001. Formation of viable but non-culturable state (VBNC) of *Aeromonas hydrophila* and its virulence in goldfish, *Carassius auratus*. *Microbiol Research*. 156: 103–106.

[21] Lloveras J. Mohamed-Issam S, Claude P, Anissa L, Philippe G. 2010. Usefulness of sputum gram stain and culture for diagnosis of pneumonia in a geriatric institution. *Journal of IMAB - Annual Proceeding (Scientific Papers)* 16: 20–22.

[22] Montalban-arques A, Peter De S, Peter B, Gregor G, Victoriano M, Delbert MG III, Jorge GV. 2015. Selective manipulation of the gut microbiota improves immune status in vertebrates. *frontiers in Immunology* , 6: 1–14.

[23] Yulvizar C. 2013. Isolasi dan Identifikasi Bakteri

Probiotik pada *Rastrelliger* sp. Isolation and Identification of Probiotic Bacteria in *Rastrelliger* sp. *Biospecies*, 6: 1–7.

[24] Logan, NA, JA Carmant, Melling, RCW Berkeley. 1985. Identification of *Bacillus anthracis* by API test. *J.Med Microbiol* 20: 75–85.

[25] Mohamadou B, CM Mbofung, G Barbier. 2013. Genotypic and phenotypic diversity among *Bacillus* species isolated from Mbuja, a Cameroonian traditional fermented condiment. *African Journal of Biotechnology* 12: 1335–1343.

[26] Amin M, Rakhisi Z, Ahmady, A.Z., 2015. Isolation and identification of *Bacillus* species from soil and evaluation of their antibacterial properties. *Avicenna J Clin Microb Infec* 2: 10–13.

[27] Ayari S, Dominic D, Moktar H, Monique L. 2016. Growth and toxigenic potential of *Bacillus cereus* during storage temperature abuse in cooked irradiated chicken rice in combination with nisin and carvacrol. *LWT - Food Science and Technology* 72: 19–25.

[28] Rengpipat S, Wannipa P, Somkiat P, Piamsak M. 1998. Effects of a probiotic bacterium on black tiger shrimp *Penaeus monodon* survival and growth. *Aquaculture* 167: 301–313.